

Control for NR-3089 (antiserum, Sheep)

Catalog No. NR-3090

This reagent is the property of the U.S. Government.

Lot (NIAID Catalog) No. G027-501-568

For research use only. Not for human use.

Contributor:

National Institutes of Allergy and Infectious Diseases (NIAID),
National Institutes of Health (NIH)

Product Description:

Reagent: Control antiserum for NR-3089

Host: Suffolk-Hampshire female yearling sheep

Immunizing Antigen: Components of crude human leukocyte interferon alpha preparations that did not bind to anti-human leukocyte interferon alpha affinity column (i.e. void volume)

NIAID Class: Research Reference Reagent

Research Reference Reagent Note (attached): No. 23

Adjuvant used: Freund's complete adjuvant in initial and booster inoculations

Material Provided/Storage:

Composition: Lyophilized

Original Volume: 0.5 mL

Storage Temperature: 4°C or colder

Reconstitution: 0.5 mL sterile distilled water

Functional Activity:

Neutralizing Titer: < 1:50 against 8 to 10 Laboratory Units of human leukocyte interferon alpha

Purity:

Sterility: No evidence of bacterial or fungal contamination

Producer and Contract:

Medical College of Pennsylvania N01-AI-82568

Citation:

Acknowledgment for publications should read "The following reagent was obtained through the NIH Biodefense and Emerging Infections Research Resources Repository, NIAID, NIH: Control for NR-3089 (antiserum, Sheep), NR-3090."

Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the following publication: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories. 5th ed.

Washington, DC: U.S. Government Printing Office, 2007; see www.cdc.gov/od/ohs/biosfty/bmb15/bmb15toc.htm.

Disclaimers:

You are authorized to use this product for research use only. It is not intended for human use.

Use of this product is subject to the terms and conditions of the BEI Resources Material Transfer Agreement (MTA). The MTA is available on our Web site at www.beiresources.org.

While BEI Resources uses reasonable efforts to include accurate and up-to-date information on this product sheet, neither ATCC® nor the U.S. Government make any warranties or representations as to its accuracy. Citations from scientific literature and patents are provided for informational purposes only. Neither ATCC® nor the U.S. Government warrants that such information has been confirmed to be accurate.

This product is sent with the condition that you are responsible for its safe storage, handling, use and disposal. ATCC® and the U.S. Government are not liable for any damages or injuries arising from receipt and/or use of this product. While reasonable effort is made to ensure authenticity and reliability of materials on deposit, the U.S. Government, ATCC®, their suppliers and contributors to BEI Resources are not liable for damages arising from the misidentification or misrepresentation of products.

Use Restrictions:

This material is distributed for internal research, non-commercial purposes only. This material, its product or its derivatives may not be distributed to third parties. Except as performed under a U.S. Government contract, individuals contemplating commercial use of the material, its products or its derivatives must contact the contributor to determine if a license is required. U.S. Government contractors may need a license before first commercial sale.

References:

1. Lowry, O. H., N. J. Rosebrough, A. L. Farr, and R. J. Randall. "Protein Measurement with the Folin Phenol Reagent." J. Biol. Chem. 193 (1951): 265-275. PubMed: 14907713.
2. Strander, H. and H. Cantell. "Production of Interferon by Human Leukocytes *In Vitro*." Ann. Med. Exp. Biol. Fenn. 44 (1966): 265-273. PubMed: 4290664.
3. Armstrong, J. A. "Semi-Micro, Dye-Binding Assay for Rabbit Interferon." Appl. Microbiol. 21 (1971): 723-725. PubMed: 4325022.
4. Havell, E. A. and J. Vilcek. "Production of High-Titered Interferon in Cultures of Human Diploid Cells." Antimicrob. Agents Chemother. 2 (1972): 476-484. PubMed: 4670440.
5. Ogburn, C. A., K. Berg, and K. Paucker. "Purification of Mouse Interferon by Affinity Chromatography on Anti-

Interferon Globulin-Sepharose." J. Immunol. 111 (1973): 1206–1218. PubMed: 4728682.

6. Berg, K., et al. "Affinity Chromatology of Human Leukocyte and Diploid Cell Interferons on Sepharose-Bound Antibodies." J. Immunol. 114 (1975): 640–644. PubMed: 1168221.

ATCC® is a trademark of the American Type Culture Collection.



NR - 3290

RESEARCH REFERENCE REAGENT NOTE # 23
CONTROL ANTISERUM (SHEEP) TO HUMAN LEUKOCYTE INTERFERON
CATALOG NUMBER G-027-501-568

Research Resources Branch
National Institute of Allergy and Infectious Diseases
National Institutes of Health
Bethesda, Maryland 20205

March, 1981

RESEARCH REFERENCE REAGENT NOTE # _____
CONTROL ANTISERUM (SHEEP) TO HUMAN LEUKOCYTE INTERFERON
CATALOG NUMBER G-027-501-568

Preparation

This serum globulin was prepared as a correlate to Sheep Antiserum to Human Leukocyte Interferon (G-026-502-568). The materials used for immunization (hereafter referred to as void) were those components of crude human leukocyte interferon (CIF) preparations (1) which do not bind to adsorbed anti-interferon globulin (2) bound to Sepharose 4B. This material is routinely recovered during the preparation of partially purified human leukocyte interferon by affinity chromatography. Materials selected for immunization showed greater than 99% reduction in antiviral activity. The rationale for using void is as follows: in addition to interferon, induced cells probably release into the growth medium numerous other cellular proteins. Selective affinity chromatography ideally retains interferon but does not remove in substantial amounts other induced or normal cell products. Immunization with this material should, therefore, yield an antiserum containing antibodies to all such products. Since the inoculum was not completely devoid of interferon, low levels of antibody to human leukocyte interferon may be present in this antiserum.

A female Suffolk yearling sheep received three intramuscular injections of 50 mg void. The first two inoculations contained Freund's Complete Adjuvant. Antiserum was collected for a five week period starting seven days after the third injection. After a six week rest, a booster injection of void with adjuvant was given, and the bleeding cycle was repeated seven days later. This pattern was followed until adequate quantities of serum were collected. The globulin portion of the serum was separated by precipitation with 50 percent ammonium sulfate, dialyzed versus 0.01 M sodium phosphate buffer pH 7, then sterilized by filtration. The globulin was dispensed (0.5 ml per ampule), freeze-dried and sealed by the American Type Culture Collection.

Recommendations for Reconstitution

Add 0.5 ml of sterile physiologic saline solution or an appropriate medium to the lyophilized powder. Precautions should be taken to avoid loss of material in the neck or stem of the ampule. The reconstituted globulin can be diluted and stored indefinitely at -20°C or lower.

Interferon Neutralization Assay

The assay procedure used at The Medical College of Pennsylvania is similar to the interferon assay in microtiter plates (3,4), except that 50 μ l volumes of serial two-fold dilutions of antiserum were preincubated for 1 hour at 37°C with 50 μ l of graded interferon dilutions covering the range from 1-32 units before addition of the 30,000 human FS-4 cells per well. Encephalomyocarditis virus at a multiplicity of 0.2 was used for challenge. Interferon, virus and cell controls were included in each test. The antiserum was titrated against several dilutions of test antigen in order to select, for computation of the titer, the series with the appropriate number of interferon units available for neutralization by antibody. The highest dilution which neutralized 8-10 reference units of interferon by partially restoring viral cytopathic effect, corrected for 1 ml volume, represented the titer of the antiserum. The human leukocyte interferon reference standard used was G-023-901-527.

Passive Hemagglutination Test

Antibody titers to three known contaminants found in the interferon preparation were measured by passive hemagglutination using the procedure of Boyden (5) as modified by Ogburn, et al. (6). Washed sheep red blood cells were treated with tannic acid (25 µg/ml) and incubated with the test antigens for 30 minutes at 37°C to facilitate binding of antigens to the surface of the red blood cells. The cells were then washed to remove unbound antigen and used as the indicator in the assay. Serial dilutions of the heat inactivated antiserum were made in a round bottom microtiter plate (Linbro Scientific #76-311-05), and 0.05 ml of a 0.6% solution of the antigen-coated sheep red blood cells was added to each well. Controls included known positive sera and antigen coated sheep red blood cells in diluent alone (PBS containing 1% normal rabbit serum). The mixture was incubated at room temperature for four hours and the pattern of hemagglutination was recorded. The presence of antibodies to the test antigen is indicated by agglutination of the antigen-coated red blood cells. The titer of the serum is defined as the reciprocal of the highest dilution showing complete hemagglutination, corrected for a one milliliter volume.

Potency

The anti-interferon titer is based on the combined results of assays performed in ten laboratories. The titers received from all laboratories were corrected to indicate a titer per milliliter for the 50 percent neutralization of 8-10 reference units of human interferon. The assigned titer of this control serum is negligible (less than 50 interferon neutralizing units per 0.5 ml).

Presumably, this antiserum contains antibody to numerous antigens of cell, virus and medium origin. Development of antibodies to three antigens known to be present (egg albumin, human albumin and an extract of human leukocyte) were monitored by passive hemagglutination throughout the immunization procedure. Antibody titers to these antigens were very similar to those found in unabsorbed antisera of a sheep immunized with partially purified human leukocyte interferon (G-026-502-568).

Results of Other Tests

Tests on randomly selected ampules showed no detectable bacterial or fungal growth. The 0.5 ml of globulin is equivalent to 17 mg protein by the Lowry procedure (7).

Use of Reference Antiserum

The purpose of this antiserum is to provide a reference reagent which can be used in conjunction with Sheep Antiserum Human Leukocyte Interferon (G-026-502-568) for the identification and characterization of biological and chemical properties attributed to human leukocyte interferon. The wide use of interferon in research has made it desirable to have standards which may be used to correlate data from different laboratories. This reagent is available in limited quantities and should be used only after preliminary studies have been performed.

The source of the reagent should be identified in each publication and a copy of all publications should be sent to the NIAID antiviral Substances Program, National Institute of Allergy & Infectious Diseases, National Institutes of Health, Bethesda, Maryland 20205.

Stability

Freeze-dried serum globulins are generally stable at room temperature (25°C) for indeterminate lengths of time. It is strongly recommended, however, that the unopened ampules be stored at +4°C or lower temperatures. The reconstituted globulin can be kept at +4°C, but a temperature of -20°C or lower is advised for long term storage.

Prepared by: Kurt Paucker, Barbara Dalton and Clifton A. Ogburn
The Medical College of Pennsylvania
Philadelphia, Pennsylvania 19129
Contract No. 1 AI 82568

References

1. Strander, H. and Cantell, K. (1966) Ann. Med. Exp. Fenn. 44:265-273.
2. Berg, K., Ogburn, C.A., Paucker, K., Mogensen, K.E. and Cantell, K. (1975) J. Immunol. 114:640-644.
3. Havell, E.A. and Vilček, J. (1972) Antimicrob. Agents and Chemother. 2:476-484.
4. Armstrong, J.A. (1971) Appl. Microbiol. 21:723-725.
5. Boyden, S.V. (1951) J. Expt. Med. 93:107-120.
6. Ogburn, C.A., Berg, K. and Paucker, K. (1973) J. Immunol. 111:1206-1218.
7. Lowry, O.H., Rosebrough, J., Farr, L. and Randell, R.J. (1951) J. Biol. Chem. 193:265-275.